

<b>Test:</b>	<b>FECAL LEUKOCYTES</b>
<b>Test ID:</b>	<b>FECL2</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Microscopic examination of stool for WBC's.
<b>Turnaround Time:</b>	Done daily 0700 - 2400. Results available the same day.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Stool.
<b>Volume:</b>	Minimum of 1 gram.
<b>Container:</b>	Clean container with a tight-fitting lid.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	a) The entire contents of the first specimen of the day is preferred. b) Avoid contamination with water, urine, paper, barium or mineral oil. Do not remove specimens from the toilet bowl. c) Specimens received in a diaper are not acceptable and will be rejected by Microbiology.
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate.

<b>Test:</b>	<b>FUNGAL SMEAR</b>
<b>Test ID:</b>	<b>FUNSO</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Microscopic exam for fungal elements by calcofluor white stain.
<b>Turnaround Time:</b>	Setup M - F. Results available the next day.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Varies.
<b>Container:</b>	Sterile container.
<b>Specimen Handling:</b>	Refer to CFUNG.

<b>Test:</b>	<b>FUNGUS CULTURE ONLY</b>
<b>Test ID:</b>	<b>CFUNO</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Culture only for yeast and molds.
<b>Turnaround Time:</b>	Done daily 0700 - 2400. Results available in 4 weeks.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Varies. See Specimen Collection instructions.
<b>Volume:</b>	Varies with specimen. See Specimen Collection instructions.
<b>Container:</b>	Sterile container, Petri dish or culturette.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	a. Respiratory: Collect according to routine sputum and bronchoscopy instructions. b. Ear, eye, nose, throat, vagina, cervix: Use culturette. c. Skin: Cleanse lesion with a 70% alcohol saturated pad. Air dry. Using a blade or blunt end of a forceps, obtain a scraping from the active border of the lesion. Place the specimen in a sterile container. d. Nails: Clean the site with a 70% alcohol saturated pad. Collect the shavings from under the nail plate into a sterile container. e. Hair: Remove dull hairs with a forceps. Place the hairs in a sterile container. f. Bone marrow: Collect in a yellow top (SPS) vacutainer tube. g. CSF: Collect 1 mL by standard aseptic technique. h. Abscess: Aspirate at least 0.5 mL into a syringe. i. Tissue: Place the specimen in a sterile container. Moisten with sterile saline. j. Urine: 20 - 50 mL of the first morning void. k. Blood: 10 mL of blood collected according to procedure for standard blood cultures.
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate urine and respiratory specimens. Store all other specimens at room temperature.

<b>Test:</b>	<b>GIARDIA ANTIGEN</b>
<b>Test ID:</b>	<b>GIARD</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Enzyme immunoassay for Giardia antigen.
<b>Turnaround Time:</b>	Done as needed. Results available in 2 - 5 days.
<b>Precollection</b>	See Ova & Parasite Exam for instructions.
<b>Instructions:</b>	
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Stool, feces-coated rectal swab. Specimens collected in diapers are acceptable for this assay.
<b>Volume:</b>	1 gram or 1 mL liquid.
<b>Container:</b>	Clean container with a tight-fitting lid.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	See Ova & Parasite Exam for instructions.
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate. Microbiology will freeze specimen upon receipt.

<b>Test:</b>	<b>GONORRHOEAE PCR</b>
<b>Test ID:</b>	<b>NGPCR</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	PCR test to detect N. gonorrhoeae. NOTE: Specimens collected using the Remel collection kit cannot be used for culture.
<b>Turnaround Time:</b>	Done daily M - F. Results available within 1-2 days.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Endocervix, vagina, penis, urethra.
<b>Volume:</b>	1 Remel swab.
<b>Container:</b>	Remel transport tube.
<b>Specimen Handling: <i>Use only Powder Free gloves.</i></b>	
<b>1. Collection:</b>	<p>a. Female:</p> <ol style="list-style-type: none"> <li>1) Remove excess mucus from the cervical os and surrounding mucosa using one of the swabs provided. Discard this swab.</li> <li>2) Insert the second swab from the collection kit 1 - 12 cm into the endocervical canal.</li> <li>3) Rotate the swab clockwise in the endocervical canal for 30 seconds to ensure adequate sampling.</li> <li>4) Withdraw the swab carefully. Avoid any contact with vaginal mucosa.</li> <li>5) Insert the swab into the Remel transport tube. Snap off the shaft at the score line or cut shaft to fit tube.</li> <li>6) Cap tube. Label with the patients name.</li> </ol> <p>b. Male:</p> <ol style="list-style-type: none"> <li>1) Insert the swab from the collection kit 2 - 4 cm into the urethra.</li> <li>2) Once inserted, rotate the swab gently clockwise, using sufficient pressure to ensure that the swab comes into contact with all urethral surfaces. Allow the swab to remain inserted for 2 - 3 seconds.</li> <li>3) Withdraw the swab. Insert the swab into the Remel transport tube.</li> <li>4) Cap tube. Label with the patients name.</li> </ol>
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate.

<b>Test:</b>	<b>GRAM STAIN</b>
<b>Test ID:</b>	<b>GSO</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Gram stain for bacteria, fungi and WBCs. Generally Gram stains are not done alone; they are included in a lower respiratory, wound, body fluid and tissue culture.
<b>Turnaround Time:</b>	Done daily 0700 - 2400. Routine: results available the same day; Urgent/STAT: within 30 minutes.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Varies.
<b>Volume:</b>	1 culturette or minimal fluid.
<b>Container:</b>	Culturette or sterile container.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	Varies. Refer to specific collection procedures.
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Varies. Refer to specific collection procedures.

<b>Test:</b>	<b>GROUP B STREP CULTURE</b>
<b>Test ID:</b>	<b>GBS</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Synonyms:</b>	Culture for Group B Streptococcus only.
<b>Turnaround Time:</b>	Done daily 0700 - 2400. Results available in 3 days.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Vagina, rectum.
<b>Volume:</b>	1 swab of each.
<b>Container:</b>	Culturette (2 swabs)
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	<p>a. With a culturette, obtain one swab each of the vaginal introitus and the anorectum.</p> <p>b. Cervical cultures are not acceptable for determining colonization.</p> <p>c. A speculum should not be used for culture collection.</p>
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate.

<b>Test:</b>	<b>H. PYLORI STOOL ANTIGEN</b>
<b>Test ID:</b>	<b>HPAG REF</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Turnaround Time:</b>	Sent to Reference Lab.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Stool
<b>Volume:</b>	1 gm
<b>Container:</b>	Sterile container
<b>Specimen Handling: Refrigerate</b>	

<b>Test:</b>	<b>HPV SCREEN, DNA PROBE</b>
<b>Test ID:</b>	<b>HPV</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	DNA probe for the detection of Human Papilloma Virus (HPV), high risk types.
<b>Turnaround Time:</b>	Results are available within 7 days.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Cervical swab, cervical biopsy, Tripath Sure Path preservative.
<b>Volume:</b>	1 swab or 5 mm biopsy.
<b>Container:</b>	Digene HPV transport tube.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	<p>a. Cervical swab: Collect specimen using the Digene swab. Insert swab into endocervical canal and rotate it in opposite directions. Rub swab over entire area of the transformation zone. Place swab in Digene transport tube. Break shaft at the prescored notch.</p> <p>b. Cervical biopsy: Immediately place the fresh biopsy (up to 5 mm in cross section) into the Digene transport tube.</p> <p>c. Preservative solutions: Follow collection device instructions.</p>
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate.

<b>Test:</b>	<b>LEGIONELLA URINE ANTIGEN</b>
<b>Test ID:</b>	<b>LUA</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Rapid immunochromatographic assay for detection of legionella pneumophila serogroup 1 antigen in urine.
<b>Turnaround Time:</b>	Done Mon – Fri, 24 hours a day, Sat and Sun 0700 – 2400. Performed Sat and Sun 0000-0700, Stat requests only. Results available the same day.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Urine (catheter or clean void).
<b>Volume:</b>	2 mL.
<b>Container:</b>	Sterile container.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	<p>a. Catheter: Swab catheter port with povidone iodine. Puncture the port with a needle and aspirate the urine into a syringe. Do NOT collect urine from the drainage bag.</p> <p>b. Clean void: Cleanse urinary meatus with towelettes. Have the patient void a small amount for discard. Collect a midstream urine specimen.</p>
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate.

<b>Test:</b>	<b>MALARIA SMEAR</b>
<b>Test ID:</b>	<b>MAL</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Microscopic examination for Malaria.
<b>Turnaround Time:</b>	Done daily 0700 - 2400. Stat requests only 0000-0700. Results available in 1 day.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Blood from a fingerstick or EDTA tube.
<b>Volume:</b>	6 thin smears and 6 thick smears on glass microscope slides.
<b>Container:</b>	Slide holder.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	<p>a. Thin smears:</p> <ol style="list-style-type: none"> <li>1) Using a 70% alcohol pad, clean the area of the finger to be punctured. Allow the skin to air dry.</li> <li>2) Puncture the finger with controlled motion using a blood lancet.</li> <li>3) Allow the blood to form a droplet by applying gentle pressure to the finger.</li> <li>4) Handling the clean slides by the edges, touch the slide to the finger and obtain a small drop of blood. Using a second clean slide, spread the blood the length of the slide. The angle between the spreading slide and the blood drop should be about 30° so that the cells are distributed evenly. Draw the film out to a feathery edge.</li> <li>5) Make 6 slides in the same manner.</li> </ol> <p>b. Thick smears:</p> <ol style="list-style-type: none"> <li>1) Using a 70% alcohol pad, clean the area of the finger to be punctured. Allow the skin to air dry.</li> <li>2) Puncture the finger with controlled motion using a blood lancet.</li> <li>3) Allow the blood to form a droplet by applying gentle pressure to the finger.</li> <li>4) Place 2-3 drops of blood on a clean slide and pool the drops into a thick film with the corner of another slide. Make the pool about the size of a dime.</li> <li>5) Allow the slide to air dry.</li> <li>6) If EDTA is used, thick and thin smears must be made within 1 hour of collection.</li> </ol>
<b>2. Transport:</b>	Deliver to Microbiology as soon as possible.
<b>3. Storage:</b>	Room temperature.

<b>Test:</b>	<b>OVA &amp; PARASITE EXAM</b>
<b>Test ID:</b>	<b>OP</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Microscopic exam (direct and concentrated smear) for intestinal parasites.
<b>Turnaround Time:</b>	Done M - F. Concentrated smear results available in 1 day. Trichrome stain results available in 7 days.
<b>Precollection Instructions:</b>	Travel history must be included. If travel history or suspect infestation are omitted, O&P will be cancelled and a Giardia Antigen test will be performed. It is imperative that all stool specimens be free of purgatives such as magnesium and oil, and materials like barium, bismuth and kaolin. Soap solutions render specimens unsatisfactory for examination. If enemas are required, only fleet or saline enemas are recommended. Specimens should not be collected for 7 - 10 days following barium or bismuth administration, and generally not for 1 - 2 weeks following antibiotic therapy like Teramycin.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Stool, bile, duodenal aspirate, ileum, sigmoid.
<b>Volume:</b>	2 grams of stool or 5 - 7 mL of liquid.
<b>Container:</b>	Clean container with a tight-fitting lid.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	<p>a. Specimen should be collected in a clean, dry container. The specimen must be free of water and urine.</p> <p>b. For best results, several specimens, preferably 3 taken at 2 - 3 day intervals, should be sent. More than one specimen from the same day should not be sent.</p> <p>c. Hard, well-formed stool is unsatisfactory for examination and will be rejected by Microbiology.</p>
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Stool: refrigerate; others: room temperature.

<b>Test:</b>	<b>PINWORM EXAM</b>
<b>Test ID:</b>	<b>PIN</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Examination of scotch tape prep for pinworms ( <i>Enterobius vermicularis</i> ).
<b>Turnaround Time:</b>	Done daily 0700 - 2400. Results available in 1 day.
<b>Precollection</b>	Patient should not be active. The specimen should be collected prior to washing or using the restroom.
<b>Instructions:</b>	using the restroom.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Scotch tape preparation of stool.
<b>Volume:</b>	1 scotch tape prep.
<b>Container:</b>	Slide holder.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	<p>a. Collect specimen in the early morning.</p> <p>b. Place a 3 inch strip of CLEAR (do NOT use frosted) scotch tape, sticky side out, over one end of a tongue depressor. Hold in place with thumb and finger.</p> <p>c. Spread buttocks and press the tongue depressor against the right and left perianal folds, being careful to cover the area between moist and dry areas.</p> <p>d. Spread the tape smoothly sticky side down on a microscope slide. Label the slide with the patient's name and date of collection. Submit the slide to Microbiology in a cardboard slide holder.</p>
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Room temperature.

<b>Test:</b>	<b>RAPID FLU</b>
<b>Test ID:</b>	<b>RFLU</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Rapid membrane immunoassay for influenza A and B. All negative flu tests will be followed with a respiratory viral culture.
<b>Turnaround Time:</b>	Done Mon- Fri 24 hours a day, 0700 – 2400 Saturday and Sunday. Results available the same day.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	NPH swab, NPH washings, nasal aspirate
<b>Volume:</b>	1 nph swab or 1ml of washings.
<b>Container:</b>	Viral Transport Media (M6) or clean container with a tight-fitting lid.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	<p><b>Nasopharyngeal Aspirate ( Best Sensitivity)</b> Suction material from the nasopharynx and place into a sterile container.</p> <p><b>Nasopharyngeal washings</b> Instruct the patient not to swallow during the procedure. With the patient's head hyperextended, instill approximately 5ml of sterile 0.85% saline into each nostril. To collect material, tilt the head forward and allow the fluid to run out of the nares into a sterile container, or aspirate the fluid by inserting a rubber bulb syringe into each nostril, then place contents into a sterile container.</p> <p><b>Nasopharyngeal Swabs</b> NOTE: Flexible Flocked NPH swab is recommended. Calcium alginate swabs CANNOT be used for this test. Carefully insert a flexible-flocked swab through the nose into the posterior nasopharynx and rotate the swab. Keep the swab near the septum and floor of the nose. Repeat the process on the other nostril. Place swab in Viral Transport Media (M6).</p> <p><b>Nasal Swabs</b> Insert a sterile swab into the nose until resistance is met at the level of the turbinates (approx. 1 inch into the nose). Rotate the swab against the nasal mucosa. Repeat the process on the other side.</p>
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients or send on ice.
<b>3. Storage:</b>	Refrigerate.

<b>Test:</b>	<b>ROTAVIRUS SCREEN</b>
<b>Test ID:</b>	<b>ROTA</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Rapid membrane immunoassay.
<b>Turnaround Time:</b>	Done daily, 0700 - 2400. Results available the same day.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Stool.
<b>Volume:</b>	1 gram.
<b>Container:</b>	Clean container with a tight-fitting lid.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	Avoid contamination with water, urine, paper or mineral oil. Specimens cannot be removed from the toilet bowl. Diapers are not accepted.
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate.

<b>Test:</b>	<b>RSV SCREEN</b>
<b>Test ID:</b>	<b>RSV</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Enzyme immunoassay (EIA) screen. If screen is negative, a culture for Respiratory Syncytial Virus (RSV) will be performed (CRV).
<b>Turnaround Time:</b>	Done Everyday, 0700 – 2400. EIA results available in 1 day. Culture results available in 3 days.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Nasal wash (specimen of choice) or NPH.
<b>Volume:</b>	Minimum of 1 mL, maximum of 2 mL of liquid or 1 NPH swab.
<b>Container:</b>	Sterile container, syringe (do NOT transport with the needle attached) or NPH swab.
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	<ul style="list-style-type: none"> <li>a. Nasal wash: Infuse 1 - 2 mL of saline into nasal passages. Use a suction device to collect washings.</li> <li>b. NPH: Flexible Flocked NPH swab is recommended. <ul style="list-style-type: none"> <li>1) Carefully insert a flexible-flocked swab through the nose into the posterior nasopharynx and rotate swab.</li> <li>2) Keep the swab near the septum and floor of the nose. Repeat the process on the other nostril.</li> <li>3) Place swab in Viral Transport Media (M6).</li> </ul> </li> </ul>
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate.

<b>Test:</b>	<b>SHIGA TOXINS 1 &amp; 2</b>
<b>Test ID:</b>	<b>SHIGA</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Shiga toxins 1 and 2.
<b>Turnaround Time:</b>	Done M– F, 0700 – 2400. Results available in 3 days.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Stool.
<b>Volume:</b>	1 gram or at least visible material on 1 culturette (2 swabs).
<b>Container:</b>	Clean container with a tight-fitting lid or 1 culturette (2 swabs).
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	<ul style="list-style-type: none"> <li>a. Stool <ul style="list-style-type: none"> <li>1) The entire contents of the first specimen of the day is preferred.</li> <li>2) Avoid contamination with water, urine, paper, barium or mineral oil. Do not remove specimens from the toilet bowl.</li> <li>3) Specimens received in a diaper are not acceptable and will be rejected by Microbiology.</li> </ul> </li> </ul>
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate.

<b>Test:</b>	<b>SHUNT PANEL</b>
<b>Test ID:</b>	<b>SHUNT</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Gram stain, Aerobic and Anaerobic Culture, Cell Count, Glucose and Protein. Sensitivity testing performed if indicated.
<b>Turnaround Time:</b>	Refer to individual tests.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Shunt or Ventricular Fluid
<b>Volume:</b>	Minimum 10 ml
<b>Container:</b>	Sterile container
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	The specimen is usually collected by the physician.
<b>2. Transport:</b>	Deliver the specimen to Microbiology immediately.
<b>3. Storage:</b>	Room temperature.

<b>Test:</b>	<b>STREP PNEUMONIAE URINE ANTIGEN</b>
<b>Test ID:</b>	<b>SPAG</b>
<b>CPT:</b>	
<b>LOINC:</b>	
<b>Test Includes:</b>	Rapid immunochromographic assay for the detection of Streptococcus pneumoniae antigen in urine.
<b>Turnaround Time:</b>	Done Mon – Fri, 24 hours a day, Sat and Sun 0700 – 2400. Performed Sat and Sun 0000-0700 Stat requests only. Results available the same day.
<b>Specimen Collection/Transport Instructions:</b>	
<b>Specimen Type:</b>	Urine (catheter or clean void).
<b>Volume:</b>	2 mL
<b>Container:</b>	Sterile Container
<b>Specimen Handling:</b>	
<b>1. Collection:</b>	a. Catheter: Swab catheter port with povidone iodine. Puncture the port with needle and aspirate urine into a syringe. Do not collect urine from a drainage bag. b. Clean Void: Cleanse urinary meatus with towelettes. Have the patient void a small amount for discard. Collect a midstream urine specimen.
<b>2. Transport:</b>	Deliver to Microbiology within 2 hours of collection for inpatients.
<b>3. Storage:</b>	Refrigerate.